

## The P-protein in *Ruta chalepensis* L.

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### Riassunto

In questo lavoro abbiamo studiato le caratteristiche della P-protein in *Ruta chalepensis* L.

Abbiamo osservato i vari tipi di P-protein già definita come nascente, tubulare e cristallina, inoltre abbiamo notato la P-protein in relazione con i microtubuli, con le spiny vescicole e con i dittiososmi.

### INTRODUCTION

Many studies on the P-protein in sieve tubes have been published. P-protein bodies consist of polymorphic aggregates showing different patterns of aggregation and extension in different species; they appear to be tubular in *Nicotiana* (CRONSHAW and ESAU, 1967) and *Coleus* (STEER and NEWCOMB, 1969), granular in *Cucurbita* (CRONSHAW and ESAU, 1968b), fibrillar in *Ricinus* (CRONSHAW, 1975), paracrystalline in certain legumes (WERGIN and NEWCOMB, 1970; ESAU, 1978).

PARTHASARATHY (1975) described various types of P-protein bodies within a single cell. Recently, FJELL (1987) studied the formation of P-protein in *Salix viminalis*.

The morphological changes of P-protein depend on differences in environmental conditions within the cell (CRONSHAW *et al.*, 1973). In a previous study we described sieve-element plastids and their ontogeny in some Rutaceae (MATARESE PALMIERI and TOMASELLO, 1983-1984).

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Key words: P-protein, Sieve-element, Rutaceae.

Lavoro eseguito con il contributo M.P.I. 60%.

In this paper, we report the characters of P-protein in *Ruta chalepensis*, with the purpose of studying the morphological changes during their ontogenesis.

#### MATERIALS AND METHODS

Excised leaves from first and second internodes of *Ruta chalepensis* L. were collected from plants growing in the Botanical Gardens of Messina, Sicily, Italy.

The samples were fixed in cold glutaraldehyde 4%, 6% buffered with 0,2M phosphate buffer at pH 7,2 for 2h, 4h, postfixed in 1% osmium tetroxide in a 0,2M phosphate buffer at pH 7,2 for 2h at 4°C, dehydrated in ethanol and finally embedded in Epon-araldite mixture (MOLLENHAUER, 1964).

The pieces were then sectioned with and Ultratome LKB microtome. The sections were stained with uranyl acetate and lead citrate (REYNOLDS, 1963) and observed with a Siemens Elmiskop 102 A electron microscope.

#### OBSERVATIONS

Various forms of P-protein bodies were observed in differentiating sieve-elements during the stages of development of leaves in *R. chalepensis*. Plastids, endoplasmic reticulum (ER), mitochondria dictyosomes, nucleus, vacuoles and principally granular P-protein are present in immature sieve-elements (Fig. 1).

During the early stages of development, we observed the disaggregation of endoplasmic reticulum; in the cytoplasm, two forms of P-protein occur tubular and nascent (Fig. 3).

Also, in longitudinal and trasverse sections, both the P-protein of tubular type (Fig. 4) and crystalline type (Fig. 2) were found.

In mature sieve-elements, where either stacks of ER in the shape of circular profiles (Fig. 7, 9) or single parietal layer appear, the P-protein is granular associated with microtubules. In the same tube the P-protein of fibrillar type occurs (Fig. 5). Fig. 2 shows coated and spiny vesicles. The companion cells show protein bodies (Fig. 6), coated vesicles with electrodense material,

dictyosomes, nucleus, a few vacuoles and many ribosomes (Fig. 6). In the sieve plates fine filaments occur.

Fig. 7 and 10 show stacks of ER in the proximity of the cell wall with circular profiles and microtubules.

Occasionally, nascent P-protein and ribosomes are close to the sieve plate in differentiating tubes (Fig. 8).

## DISCUSSION

Our present observations were undertaken with the purpose of completing our previous study in some Rutaceae on sieve-element plastids (MATARESE PALMIERI and TOMASELLO, 1983-1984).

The occurrence of P-protein was briefly reported and commented; in fact in *Calodendrum capensis* the P-protein is of fibrillar type, in *Citrus limon* of fibrillar type, in *R. chalepensis* of fibrillar, tubular and crystalline type, but the origin was not discussed.

During early stages of development, either the sieve-elements, the companion cells, or the near parenchymatous cells, show nascent, tubular and crystalline forms of P-protein. The P-protein described by ESAU and CRONSHAW (1967) was studied as an amorphous, fibrillar, granular, tubular and crystalline type. During early stages of development of *Ruta*, the P-protein consists of fibrills associated with tubules as observed by CRONSHAW and ESAU (1968 a, b) and ESAU (1971). We reached the same results as the above mentioned Authors; in fact the P-protein in at first of nascent type and finally of crystalline type.

Our study in *Ruta*, therefore, also agrees with Fjell's observations (1987), who observed the ontogeny of P-protein in *Salix viminalis*: the nascent and tubular P-protein. In some other Rutaceae we observed the tubular and fibrillar type P-protein but not the granular P-protein (MATARESE PALMIERI and TOMASELLO, 1983-1984). In some Aurantioideae we observed only the fibrillar P-protein because we did not investigate the ontogeny of P-protein (MATARESE PALMIERI and TOMASELLO, 1983-1984). According to HOEFERT (1979-1980), coated vesicles are involved in the formation of granular P-protein. CRONSHAW and ESAU (1968 a, b) described the occurrence of vesicles with electron-dense material close

to the P-protein and considered them, as « spiny vesicles ». Also NEWCOMB (1967) observed the spiny vesicles during the formation of P-protein.

We described the occurrence of vesicles in some Rutaceae; now the present observations in *R. chalepensis* showed the vesicles close to dictyosomes with electron-dense material and closed to P-protein. On these basis, as in HOEFERT (1979) and CRONSHAW and ESAU (1968 a, b), the proximity of dictyosomes to the coated vesicles and to P-protein suggests that the P-protein takes origin from dictyosomes. The P-protein could rise from vesicles with a rupture followed by a discharge of their contents as interpreted by CRONSHAW and ESAU (1968 a, b). The spiny vesicles are closed to the nascent P-protein, while the coated vesicles seem to merge with P-protein bodies.

#### ACKNOWLEDGEMENTS

I would like to thank Prof. Giacomo Tripodi for reading the manuscript.

#### SUMMARY

In this paper we report the characters of the P-protein in *Ruta chalepensis* L. We observed the nascent, tubular and crystalline type of P-protein. Also we observed the P-protein associated with microtubules, with spiny vesicles and dictyosomes.

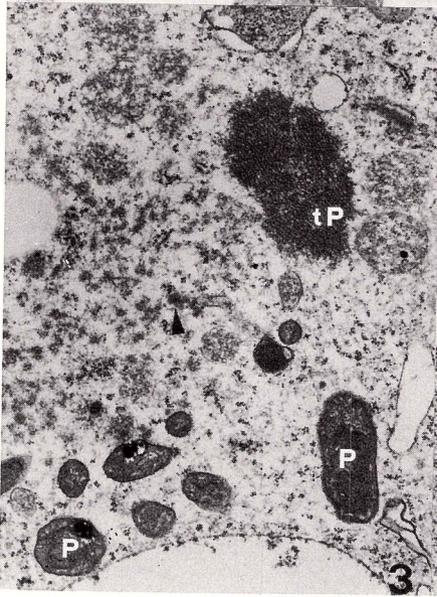
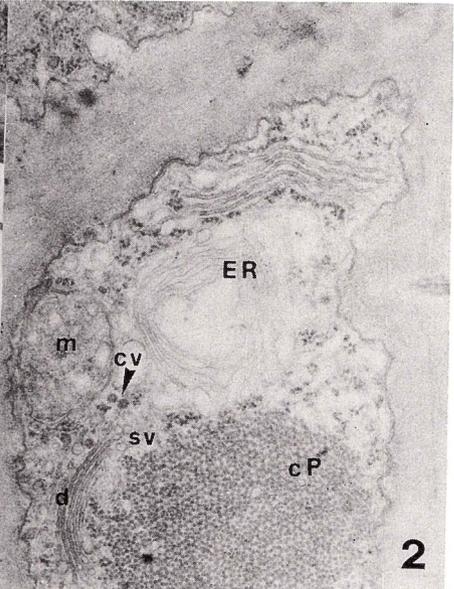
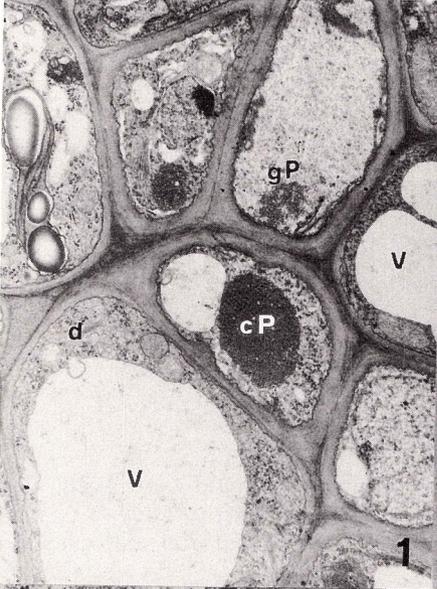
#### REFERENCES

- CRONSHAW J., 1975. *P-proteins*. In: Aronoff S., Dainty J., Gorham PR, Srivastava LM, Swanson CA (eds). *Phloem transport*. NATO Ady Study Inst Ser, Plenum Press. New York London, pp. 79-147.
- CRONSHAW J., ESAU K., 1967. *Tubular and fibrillar components of mature and differentiating sieve elements*. J. Cell. Biol., 34: 801-816.
- CRONSHAW J., ESAU K., 1968 a. *P-protein in the phloem of Cucurbita. I. The development of P-protein bodies*. J. Cell. Biol., 38: 25-39.
- CRONSHAW J., ESAU K., 1968b. *P-protein in the phloem of Cucurbita. II. The P-protein of mature sieve elements*. J. Cell. Biol., 38: 292-303.
- CRONSHAW J., GILDER J., STONE D., 1973. *Fine structural studies of P-proteins in Cucurbita, Cucumis and Nicotiana*. J. Ultrastruct. Res., 45: 192-205.

- ESAU K., 1971. *Development of P-protein in sieve elements of Mimosa pudica*. *Protoplasma* 73: 225-238.
- ESAU K., 1978. *Developmental features of the primary phloem in Phaseolus vulgaris L.* *Ann. Bot.*, 42: 1-13.
- ESAU K., CRONSHAW J., 1967. *Tubular components in cells of healthy and tobacco mosaic virus-infected Nicotiana*. *Virology*, 33: 26-35.
- FJELL I., 1987. *P-protein and inclusion bodies in root protophloem of Salix viminalis*. *J. Bot.*, 7: 305-310.
- HOEFERT L.L., 1979 a. *Ultrastructure of developing sieve elements in Thlaspi arvense L. I. The immature state*. *Amer. J. Bot.*, 66: 925-932.
- HOEFERT L.L., 1980 b. *Ultrastructure of developing sieve elements in Thlaspi arvense L. II Maturation*. *Amer. J. Bot.*, 67 (2): 194-201.
- MATARESE PALMIERI R. and TOMASELLO D., 1983-84. *Sieve element plastids in some species of the Rutaceae*. *Delpinoa n.s.*, 25-26: 29-37.
- MOLLENHAUER H.H., 1964. *Plastic embedding mixtures for use in electron microscopy - Stain*. *Technol.*, 39: 111-114.
- NEWCOMB E.H., 1967. *A spiny vesicle in slime-producing cells of the bean root*. *J. Cell. Biol.*, 35: 17-22.
- PARTHASARATHY M.V., 1975. *Sieve element structure*. In: Zimmermann MH, Milburn JA (eds). *Transport in plants. I. Phloem transport*. *Encyclopedia of plant physiology, new series Vol. 1*. Springer, Berlin Heidelberg New York, pp. 3-38.
- REYNOLDS E.S., 1963. *The use of lead citrate at a high pH as an electron opaque stain in electron microscopy*. *J. Cell. Biol.*, 17: 208-212.
- STEER M.W., NEWCOMB E.F., 1969. *Development and dispersal of P-protein in the phloem of Coleus blumei Benth.* *J. Cell. Sci.*, 4: 155-169.
- WERGIN W.P., NEWCOMB E.H., 1970. *Formation and dispersal of crystalline P-protein in sieve elements of soybean (Glycine max L.)* *Protoplasma*, 71: 365-388.

Differentiating sieve-elements of *R. chalepensis*.

- Fig. 1. - Cross section of a leaf fixed in glutaraldehyde 4% for 2h. Differentiating phloem with dictyosomes (d), vacuoles (V), granular (gP) and crystalline (cP) P-protein. X27000.
- Fig. 2. - Cross section of a leaf fixed in glutaraldehyde 6% for 4h. Note the changes of endoplasmic reticulum (ER), presence of mitochondria (m), coated vesicle (cv), spiny vesicles (sv), dictyosomes (d), polisomes and the P-protein of crystalline-type (cP). X32000. (From Matarese Palmieri and Tomasello, 1983-1984).
- Fig. 3. - Cross section of a leaf fixed in glutaraldehyde 6% for 4h. Presence of tubular (tP) and nascent (see arrow) P-protein, plastids (P). X18000.
- Fig. 4. - Longitudinal section of a leaf fixed in glutaraldehyde 6% for 4h with the Pprotein of tubular type (tP). X14000.



Differentiating sieve-elements of *R. chalepensis*.

- Fig. 5. - Cross section of a leaf fixed in glutaraldehyde 4% for 2h. The P-protein granular (gP) associated with microtubules (mt), presence of fibrillar P-protein type (fP) and parietal endoplasmic reticulum (ER). X32000.
- Fig. 6. - Cross section of a leaf fixed in glutaraldehyde 4% for 2h. Companion cell shows granular protein body (gP) coated vesicles (cv) dictyosomes (d), nucleus (N) and ribosomes (r). X33000.



Differentiating and mature sieve-tube.

- Fig. 7. - Cross section of a leaf fixed in glutaraldehyde 4% for 2h. Presence in mature sieve-tube of ER circular profiles, and microtubules (mt). X18000.
- Fig. 8. - Section of leaf fixed in glutaraldehyde 6% for 2h with the sieve plate differentiating, note the nascent P-protein (see arrow). X18000.
- Fig. 9. - Leaf fixed in glutaraldehyde 4% for 2h. Observe the circular profiles of ER and the P-protein in sieve tube (Pp). X18000.
- Fig. 10. - Leaf fixed in glutaraldehyde 4% for 2h. Note fine filaments of P-protein in sieve plate (fP). X36000.

